

Comparison between a Clostridium tyrobutyricum - specific quantitative polymerase chain reaction (qPCR) method and a traditional method for determining total spore-forming bacteria in clover-grass silage

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Introduction

- The most probable number (MPN) method is commonly used to estimate total spore counts (Jonsson 1990).
- MPN method is time consuming and not specific for C. tyrobutyricum.

The qPCR method can quickly quantify and identify microorganisms by their DNA

sequences, such as C. tyrobutyricum.



Cheese made from milk with high count of *C. tyrobutyricum*



Objectives

• To investigate potential differences and relationships in the clostridia estimate of silage between the qPCR method and the MPN method.

- To evaluate relationships between silage fermentation characteristics and
- 1) *C. tyrobutyricum* copies (qPCR)
- 2) total spore-forming bacteria count (MPN)



Material and Methods



Sward of 75% red clover/25% grass Harvest September 4, 2011 in third cut at Skara, southwest Sweden



Wilted to 28-35% DM 16% crude protein of DM 10% WSC of DM



Harvest

Inoculants applied at chopper

Homofermentative LAB
Heterofermentative LAB
Homo/Hetero LAB
Control without inoculant
14 silage jars stored for 142 d







Methods for clostridia estimate

qPCR method TaqMan

Fast DNA kit for soil and lysozyme to disrupt cell walls

The Probe, forward- and reverse primer used (Bassi et al., 2013)

Primer/probe	Sequence
Primer forward (ptaF)	5'- AAG GGA AGT GCA CAA CAT GA - 3'
Primer reverse (ptaR)	5'- ACT ACC AGG TGC TTT TAA ATT TGC - 3'
Probe (ptaP) 5'- FAM -	TGC TAA ACA AAT GCA ACC AGA – TAMRA - 3'

Number of gene copies/g silage was calculated

MPN

Bryant Burkey broth (BBB) as substrate, 37°C for 7 days





Statistical analysis

- Least-square (LS) means between methods were compared in PROC GLM of SAS
- Linear regression analysis in PROC REG of SAS to investigate relationships between total spore forming bacteria count by the MPN method and the *C.* tyrobutyricum numbers by the qPCR method
- Stepwise regressions in PROC REG of SAS to investigate relationships between silage fermentation characteristics and
- 1) *C. tyrobutyricum* by qPCR
- total spore count by MPN
- P < 0.15 as the significance level for inclusion of a variable in the model



Results

Composition	Range of 14 samples		
C. tyrobutyricum, log gene copies g ⁻¹	2.7 – 7.0		
Total spore-forming bacteria, log MPN g ⁻¹	0.9 - 7.0		
pH 3 days	4.38 – 5.18		
Final pH	4.19 – 4.94		
Lactic acid, % DM	3.0 – 11.3		
Acetic acid, % DM	2.0 – 4.9		
Propionic acid, % DM	0.00 - 0.96		
Butyric acid, % DM	0.00 – 1.05		
Ethanol, % DM	0.24 - 0.78		
1,2 - Propandiol, % DM	0.03 - 0.43		
NH ₃ -N, % of total N	8.0 – 15.1		
WSC, % of DM	0.31 – 0.81		



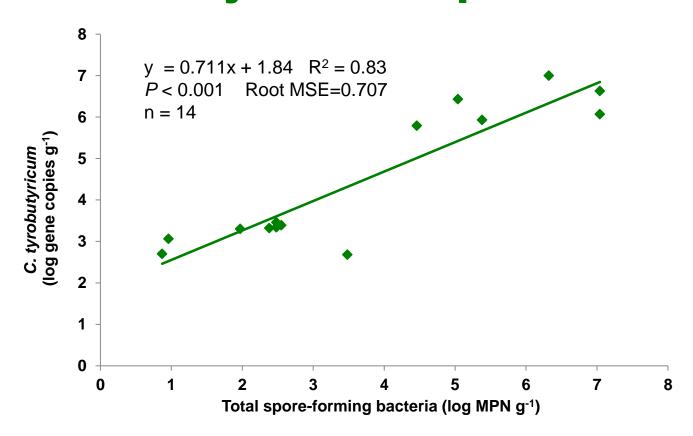
Comparison between methods

LS means for each method n = 14 silage samples

MPN	qPCR	SEM	P
3.7 log MPN g ⁻¹	4.5 log copies g ⁻¹	0.51	0.301



Relationship between clostridia estimates by MPN and qPCR





Relationships between fermentation characteristics and *C. tyrobutyricum* and total spore-forming bacteria

Method	Fermentation parameter	P	Slope	Root MSE	R ²
qPCR	pH 3 days	< 0.001	+		
	Lactic acid	< 0.001	-		
	Acetic acid	< 0.05	+		
	1,2 - Propanediol	0.107	-	0.239	0.98
MPN	Butyric acid	< 0.001	+		
	Butanol	0.113	+	0.719	0.90



Conclusions

- The qPCR method for specific detection of *C. tyrobutyricum* in clover-grass silage seems promising but needs to be investigated further in comparison to the MPN method.
- Using a larger data set, more robust relationships with the fermentation products can be developed.



THANK YOU FOR YOUR ATTENTION!!



